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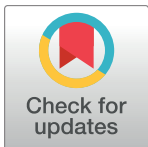
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RESEARCH ARTICLE

The responses of an anaerobic microorganism, *Yersinia intermedia* MASE-LG-1 to individual and combined simulated Martian stresses

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Abstract

The limits of life of aerobic microorganisms are well understood, but the responses of anaerobic microorganisms to individual and combined extreme stressors are less well known. Motivated by an interest in understanding the survivability of anaerobic microorganisms under Martian conditions, we investigated the responses of a new isolate, *Yersinia intermedia* MASE-LG-1 to individual and combined stresses associated with the Martian surface. This organism belongs to an adaptable and persistent genus of anaerobic microorganisms found in many environments worldwide. The effects of desiccation, low pressure, ionizing radiation, varying temperature, osmotic pressure, and oxidizing chemical compounds were investigated. The strain showed a high tolerance to desiccation, with a decline of survivability by four orders of magnitude during a storage time of 85 days. Exposure to X-rays resulted in dose-dependent inactivation for exposure up to 600 Gy while applied doses above 750 Gy led to complete inactivation. The effects of the combination of desiccation and irradiation were additive and the survivability was influenced by the order in which they were imposed. Ionizing irradiation and subsequent desiccation was more deleterious than vice versa. By contrast, the presence of perchlorates was not found to significantly affect the survival of the *Yersinia* strain after ionizing radiation. These data show that the organism has the capacity to survive and grow in physical and chemical stresses, imposed individually or in combination

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that are associated with Martian environment. Eventually it lost its viability showing that many of the most adaptable anaerobic organisms on Earth would be killed on Mars today.

Introduction

Considering the planetary bodies in our solar system in terms of their environmental conditions, Mars is the most Earth-like planet, at least in its early history [1–3]. A crucial question remains as to whether habitable, potentially inhabited, environments exist or have existed on Mars. Assessing the potential habitability of Mars and detecting possible life, depends on knowledge of whether combined environmental stresses experienced on Mars are compatible with life as we know it and whether such a record of life could ever be detected. Our current ability to make these assessments is hampered by a lack of knowledge of how combined effects of different environmental stresses influence survival and growth of microorganisms. A range of physical extremes can be examined to assess whether Martian environments have been or are habitable. These stresses are: low temperature (in surface and subsurface environments), high salinity and oxidizing compounds (caused by combinations of salts on Mars, including chlorides, sulfates, perchlorates and the presence of oxidants in soil), aridity (during periods of transient water activity either in the past or present), low availability of nutrients (possibly in all locations), high ionizing radiation (particularly in surface environments on Mars, although these stresses would be less important for deep subsurface life) and anoxic conditions in all Martian environments [3–5]. Some of these factors (e.g. perchlorate and UV-radiation) even strengthen each other mutually in terms of hostility to life [6].

Past attempts to investigate the response of organisms to Martian extremes have generally focused on aerobic organisms including spores of *Bacillus subtilis*, vegetative aerobic halophiles and some phototrophs [7–10]. However, Mars is an anoxic planet and a question that remains unanswered is how anaerobic microorganisms survive under stresses associated with present-day Martian conditions. To address this question, we need to subject highly adaptable anaerobic microorganisms found in diverse environments on the Earth to Martian conditions to ascertain whether characteristics that allow for persistence in extreme physicochemical conditions on Earth would allow these organisms to survive on Mars. This is especially the case for microorganisms that have been accidentally deposited via robotic missions on Mars known as forward contamination. In the context of planetary protection, especially in the case of a mission to Mars, the microbial communities residing in spacecraft assembly clean rooms and on the spacecraft itself were analyzed and their tolerance to Martian conditions was tested. However, their focus was mostly laid on aerobes and single stresses [11–15].

This paper describes the stress response of *Yersinia intermedia* MASE-LG-1 a facultatively anaerobic organism isolated from an extreme Icelandic environment in the framework of MASE (Mars Analogues for Space Exploration) [16]. The following stresses relevant to the Martian environment were tested to advance our knowledge of survivability of a facultatively anaerobic model organism: desiccation, ionizing radiation (X-rays), low pressure, simulated Martian atmosphere, salts and oxidizing compounds, high / low pH, and temperature. In particular, we were interested in the effects of combined environmental stresses, for which data on microbial responses are poorly developed or absent. We report these results and discuss implications for the survival of known anaerobic microorganisms on Mars.

This study will advance our general understanding of the conditions for anaerobic microorganisms on Mars and has implications for the survival of microorganisms accidentally deposited on Mars in future robotic or human missions and whether they would survive.

Materials and methods

Sampling, enrichment and purification

The sampling site, Lake Grænavatn (63° 53.07' N, 22° 3.70' W, Iceland), is a restricted area and the sampling campaign was organized and accompanied by the MASE-cooperation partner Viggo T. Marteinsson (MATIS—Prokaria, Reykjavík, Iceland). No protected species were sampled.

Y. intermedia MASE-LG-1 was isolated from this Icelandic field site (Lake Grænavatn). The lake is a maar type explosion crater, about 35 km south west of Reykjavík in the Reykjanesfólkvangur National Park [17]. Lake Grænavatn is about 45 m deep, approximately 360 m × 260 m in size and hydrothermal activity can be found on the bottom of the lake [17]. The water temperature at the sampling sites were approximately 4°C with a pH-value of around 2. To maintain an oxygen free environment, sampling was performed under anoxic conditions. A determination of the oxygen level at the sampling site was not performed. The samples were taken from the waterside of the lake with a sterile scoop attached to the end of a 1.25 m long pole. Thus, a depth of 30 to 40 cm below the water level was reachable. Water and aqueous / sediment samples were transferred in 100 ml Duran glass flasks. To maintain an anoxic environment, the samples were immediately reduced by injecting a reducing agent (1 ml cysteine-HCl of a 2.5% (w/v) solution) and sealed gastight. Subsequently, microbial enrichments were obtained using the specifically designed MASE I medium, a minimal medium with the addition of various supplements.

MASE I medium contains per liter: NH_4Cl 0.5 g, NaHCO_3 0.2 g, NaH_2PO_4 0.06 g, 10 x Wolfe's minerals 1 ml [18], 10 x Wolfe's vitamins 1 ml [16], cysteine-HCl 0.5 g. The medium was portioned (20 ml each) into 120 ml serum bottles and gassed with 1 bar N_2/CO_2 (80/20 vol/vol) [19]. Prior to inoculation, the medium was supplemented with 0.01% (w/v) KNO_3 , 0.01% (w/v) C-Org-Mix (C-Org-Mix stock solution contains per 100 ml: yeast extract 1 g, peptone 1 g, brain heart infusion 1 g, meat extract 1 g, 10 x Wolfe's Vitamins 100 µl). The pH of the MASE I medium was adjusted to 7.0. The incubation was carried out at 30°C and on a shaker at 50 rpm. Strain MASE-LG-1 was enriched and purified in MASE I medium.

Microscopic observations of the microbial isolate

For electron microscopy, a fresh culture was fixed in paraformaldehyde (final concentration 2.5%), washed three times for 15 min in sodium phosphate buffer (pH 7.4) and filtered onto a 0.2 µm filter. For scanning electron microscopy (SEM), filters were placed into microporous ceramic capsules for dehydration using increasing ethanol / distilled water solutions (25%, 50%, 75% ethanol 15 min each, and 100% ethanol 1 h) followed by a series of ethanol / acetone treatments (25%, 50%, 75% acetone 15 min each, and 100% acetone 1 h). Samples were gold coated and finally observed and analyzed with a Hitachi S4500 Field Emission Gun Scanning Electron Microscope, equipped with an EDX detector (Oxford Instruments). For transmission electron microscopy (TEM), contrast was increased by osmium tetroxide coloration using an OsO_4 4% aqueous solution. The upper part of the filter was gold-coated prior to dehydration in order to distinguish the upper and the lower parts of the filters during TEM observations. Dehydration baths for TEM and SEM were identical. For ultrathin sectioning, the samples were prepared by embedding in TAAB 812 Resin (TAAB Laboratories), with a series of acetone / resin mixtures in the proportions 3:1, 1:1, and 3:1, followed by one bath in 100% resin (overnight). The resin blocs were finally left to harden at 60°C in a last 100% resin bath. Ultrathin sections were made using a diatom diamond knife mounted on a Reichert ultra-microtome and placed on copper TEM grids. Observations and analysis were made with a Philips CM20 Transmission Electron Microscope, equipped with an EDX detector.

Taxonomic classification of the microbial isolate

For phylogenetic identification, gDNA was isolated by the XS buffer method [20]. DNA was amplified by PCR using a 16S rRNA gene primer pair: forward 9Bf and reverse 1406uR [21, 22]. PCR products were purified (PCR purification kit, Quiagen) and Sanger sequenced (GenArt®). Subsequently, the sequence was submitted to GenBank using Sequin 12.30 (www.ncbi.nlm.nih.gov/Sequin/). The sequence is publicly available (GenBank accession number KX964607). Phylogenetic classification was assessed via BioEdit [23], MEGA6 [24] and MALDI-TOF (following protocol 3 as described previously [25]). To identify the strain at species level two physiological test series, both inoculated with a fresh culture and prepared according to the manufacturer's instruction were applied (Api® 20A, Rapid ID 32A; bioMérieux).

Determination of optimal growth conditions

All growth experiments were carried out in triplicates and microbial growth was determined by measuring changes in the optical density at 600 nm. Tested temperatures were 4°C, 25°C, 30°C, 35°C, and 40°C. The pH tests (pH 3–pH 9 at increments of pH 1) were carried out at optimal growth temperature (T_{opt} : 30°C). Sterile anoxic stock solutions of 2 M HCl (acidic pH) and 2 M NaOH (alkaline pH) were used to adjust the pH of the medium. At optimal growth conditions, a growth curve was determined by direct cell counting using a Thoma counting chamber.

Stress tests and determination of the survival rate

Determination of the survival rate and enumeration of cultivable cells was achieved by the most probable number (MPN) technique via dilution series with ten-fold dilution steps [26]. If not indicated differently, MPN dilution series and stress tests were performed with cells grown in anoxic MASE I medium (Table 1). All experiments were repeated independently at least three times, representing biological replicas. The plotted data represent mean values with

Table 1. Overview of performed single and combined stress tests with strain MASE-LG-1.

Single stress	Conditions
Desiccation	≤ 6 month
X-rays	≤ 1000 Gy
Nutrient limitation	Dilution of medium and nutrients (1:10 / 1:50)
Perchlorates	15 min, ≤ 3 M
Osmotic stress (NaCl)	15 min, ≤ 6 M
H ₂ O ₂	15 min, ≤ 1 M
Temperature	-86°C—+90°C
Combined stresses	Conditions
Desiccation and vacuum	≤ 1 month
Desiccation and Mars atmosphere	≤ 1 month
Desiccation and X-rays	1 day; ≤ 800 Gy
X-rays and oxygen	≤ 800 Gy; 21% O ₂
X-rays and desiccation	≤ 800 Gy; 1 day
Nutrient limitation and desiccation	1:10 / 1:50; 1 day
Nutrient limitation and X-rays	1:10 / 1:50; ≤ 600 Gy
Perchlorate and desiccation	0.5% / 1%; 1 day
Perchlorate and X-rays	0.5% / 1%; ≤ 700 Gy
NaCl and desiccation	≤ 6 M (15 min); 1 day

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standard deviations. Significance was tested by ANOVA analysis of variance. *P*-values < 0.05 were considered statistically significant and were marked by an asterisk within the figures.

The survival rate (*S*) was calculated as relative survival after cell damaging treatment (*N*) compared to the non-treated control (*N*₀) (*S* = *N*/*N*₀). *D*₁₀-values were only determined after irradiation. The *D*₁₀-values are indicating the dosage in Gy which reduces the survival rate by one order of magnitude were calculated from the regression lines of the exponential slopes of the survival curves [27].

Desiccation experiments and exposure to vacuum or Martian atmosphere. Cells were cultivated under optimal growth conditions until stationary growth phase was reached (~ 24 h). Desiccation experiments were performed as described earlier [28]. One milliliter of culture (equivalent to 10⁷ cells) was spread evenly on sterile glass slides or quartz discs and dried under oxic laboratory conditions (room temperature, relative humidity 33 ± 3.5%) or under anoxic conditions in an anaerobic chamber (Coy Laboratory Products Inc.; [O₂] < 5 ppm, relative humidity 13 ± 0.5%).

Exposure to vacuum or Martian atmosphere was carried out using a newly designed transport and exposure box (Trex-Box) (Fig 1). The Trex-Box is a gastight closable stainless steel box (13.5 cm long, 13.5 cm wide, 5.0 cm high) with one borehole where various pumping systems can be connected which allows for the regulation of the conditions inside the box. The box contains three layers of sample carriers. Each sample carrier has space enough for 16 separate samples (Fig 1A). Desiccated cells on quartz discs were transferred into the Trex-Box under anoxic conditions and were exposed subsequently to low pressure or ionizing radiation.

High vacuum (5 × 10⁻⁵ Pa) was produced by a pumping system (Rotary vane pump DUO 035 and HIPace 700; Pfeiffer Vacuum GmbH) and continuously monitored (TPG 262 Full Range Gauge; Pfeiffer Vacuum GmbH) during exposure. For simulating the Martian atmosphere, a Mars-like gas composition (2.7% N₂, 1.6% Ar, 0.15% O₂ in CO₂ vol/vol) was used at a pressure of 10³ Pa (Fig 1B).

For exposure to ionizing radiation, desiccated cells on glass slides were transferred into gas-tight closable glass vessels which are normally used for HPLC measurements. Subsequently, the dried cells on glass slides were exposed to X-rays under anoxic conditions.

Irradiation experiments. Exposure to ionizing radiation was carried out as described in earlier studies [29]. Cell cultures (in liquid suspension) in HPLC vessels or desiccated cells on glass slides were used. Irradiation was carried out with the X-ray source Gulmay RS 225A (Gulmay Medical Ltd.) at 200 kV and 15 mA. The cells were irradiated at a distance of 19.5 cm below the X-ray source with 20 Gy/min ± 5 Gy/min up to 1000 Gy. The dose rate was

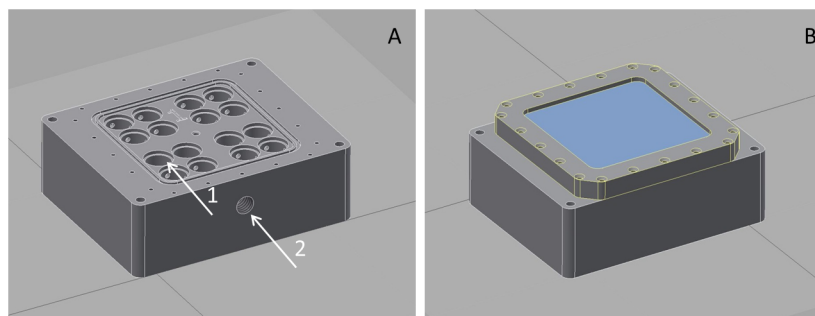


Fig 1. The Trex-Box. The desiccated cells on quartz glass discs can be positioned (arrow 1) inside the open Trex-Box. The box can be connected to various pumping systems (arrow 2) (A). Closed Trex-Box is ready for exposure to combined stresses e.g. radiation and vacuum (B).

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measured with a UNIDOS dosimeter (PTW Freiburg GmbH). All irradiation experiments were performed at room temperature.

Exposure to Mars relevant chemical compounds. Different concentrations of salts (NaCl, MgCl₂, CaCl₂), Martian relevant concentrations of perchlorates (NaClO₄, Mg(ClO₄)₂, Ca(ClO₄)₂ [30]) and hydrogen peroxide (H₂O₂) were added to an overnight culture grown at optimal conditions (Table 1). After 15 minutes of incubation to these compounds, exposure was terminated by an experimentally feasible high dilution step (1:400) and the MPN assay was applied to determine the survival rate.

Exposure to different levels of nutrient availability. To test if the cells are able to grow under nutrient limitation, they were cultivated in medium with a lower content of ingredients (diluted medium). All salts and added supplements were scaled down to 1:10 or 1:50 compared to standard cultivation medium.

Combined stresses. To investigate whether stresses act in a synergistic, antagonistic or additive way, several combinations of single stresses were tested (Table 1). To avoid any cellular repair effects, exposure to the combined stressors was done in the shortest time possible.

Results

Six different bacterial strains, growing under organism specific conditions, were isolated from Lake Gränavatn [16]. One of the strains, namely MASE-LG-1 will be described in detail.

Strain MASE-LG-1 is affiliated to the genus *Yersinia*

Strain MASE-LG-1 is a motile, rod-shaped, gram negative microorganism $\leq 1.5 \mu\text{m}$ long (Fig 2A and 2B), growing to a maximal cell density of approximately 10^7 cells per ml in MASE I medium (Fig 2C).

16S rDNA gene sequence and MALDI-TOF (data not shown) analyses indicated that strain MASE-LG-1 is affiliated to the bacterial genus *Yersinia* (phylum Proteobacteria; S1 Fig). The closest relatives were *Y. intermedia*^T DSM 18517 [31] and *Yersinia aldovae*^T DSM 18303 [32]. Biochemical identifications (API and Rapid test; S1 Table) identified the isolate MASE-LG-1 as *Y. intermedia*. Isolation of putative plasmids revealed that *Y. intermedia* MASE-LG-1 does not possess any plasmid(s).

Y. intermedia MASE-LG-1 has been deposited at the Leibniz Institute DSMZ-German Collection of Microorganisms and Cell Cultures and is listed under the DSMZ Acc. No. DSM 102845.

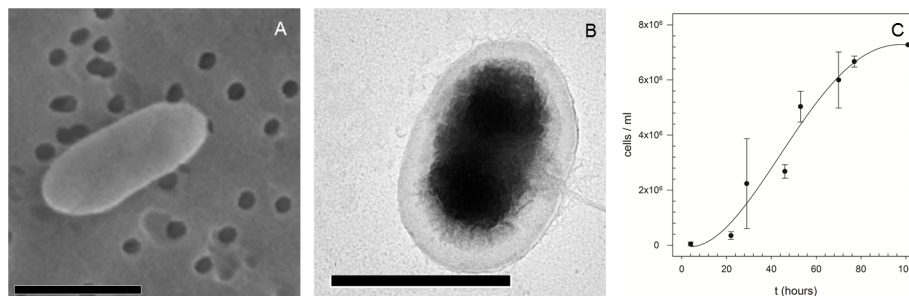


Fig 2. Microscopic images of *Y. intermedia* MASE-LG-1. SEM (A) and TEM (B) images of *Y. intermedia* MASE-LG-1 and growth curve (C) of *Y. intermedia* MASE-LG-1 cultivated under optimal anoxic conditions (30°C, pH 7). Growth was determined by direct cell counting using a Thoma counting chamber (n = 3). Bars: 1.0 μm .

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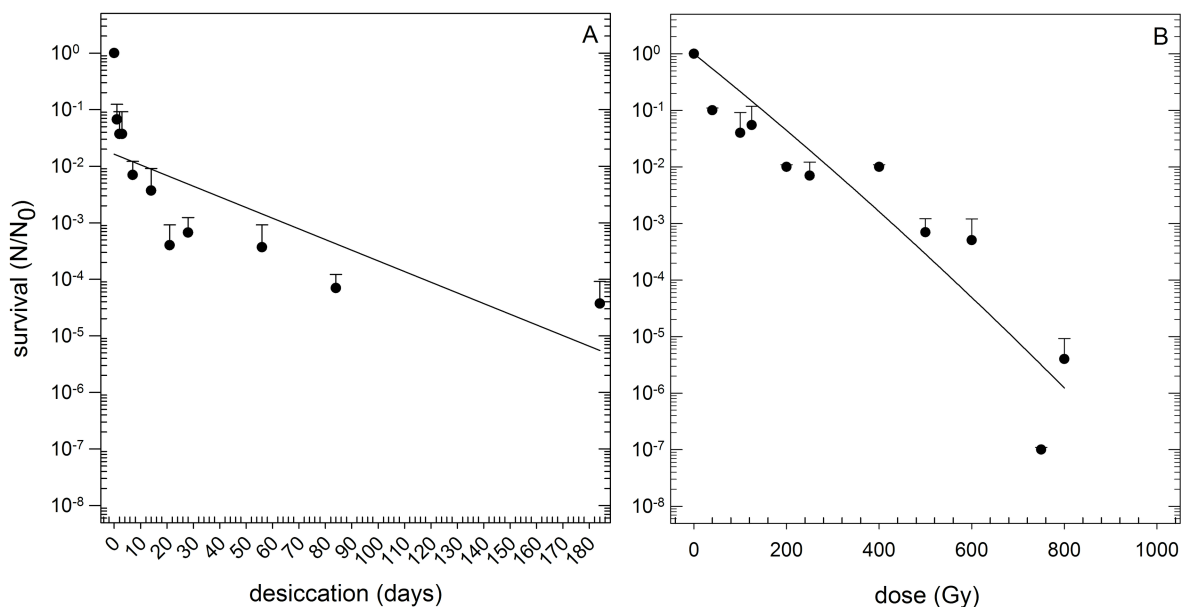


Fig 3. Survival after desiccation and radiation. Survival of *Y. intermedia* MASE-LG-1 after anoxic desiccation (A) and after exposure to ionizing radiation (B). N₀: Viable cells without desiccation / irradiation, N: Viable cells after desiccation / irradiation (n = 3). (A) Cells were applied on glass slides and dried under anoxic conditions up to 190 days. (B) Cells were exposed to ionizing radiation up to 1000 Gy in liquid culture medium under anoxic conditions.

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Evaluation of optimum growth conditions

The temperature range for growth was found to be 25°C–35°C (optimum of 30°C). No growth occurred at 4°C and above 40°C. The pH range for growth was 5–8 with an optimum of pH 7. Under optimal conditions a generation time of 5 hours and 18 minutes was determined (Fig 2C).

Y. intermedia MASE-LG-1 survives prolonged periods of water loss and exposure to ionizing radiation

Y. intermedia MASE-LG-1 possesses the ability to survive prolonged periods of desiccation under anoxic conditions. The survival rate decreased rapidly after one day of desiccation ($S(24\text{ h}) = 6.7 \times 10^{-2}$, Fig 3A). After the first 24 h, the survival rate declined more slowly to approximately one order of magnitude per month. After three months of desiccation, the survival was reduced by four orders of magnitude ($S(84\text{ days}) = 7 \times 10^{-5}$). Cells dried under anoxic conditions were exposed to vacuum representing an extreme form of desiccation and Martian atmosphere. Exposure to vacuum (10⁻⁵ Pa) and to Martian atmosphere (Mars gas at a pressure of 10⁻³ Pa) led to an additional reduction of survival of one to two orders of magnitude lower than that observed after anoxic desiccation under one terrestrial atmosphere (Fig 4).

By contrast, the strain was sensitive to ionizing radiation. After exposure to 150 Gy, detrimental effects on survivability were visible leading to an overall reduction of more than one order of magnitude (Fig 3B). After exposure to 750 Gy, only marginal survival was observed and after exposure to 1000 Gy, no viable cells were detectable. The calculated D₁₀-value was 190 Gy.

Additive effects of desiccation and ionizing radiation

In general, the combination of irradiation and desiccation led to additive effects (Fig 5A). However, the manifestation of the additive effect was influenced by the order of the stress

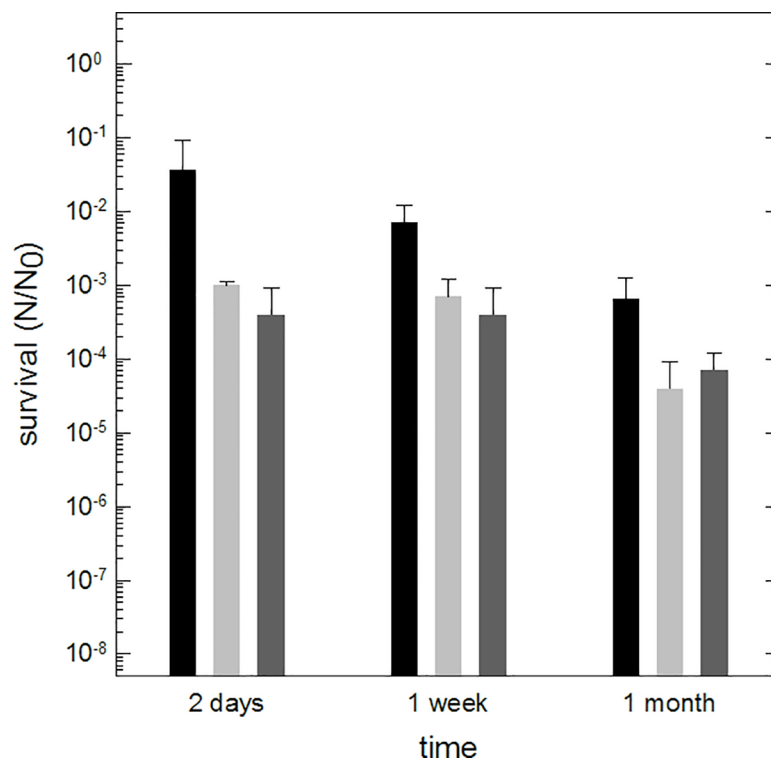


Fig 4. Survival of *Y. intermedia* MASE-LG-1 after exposure to desiccation, vacuum and Martian atmosphere. N_0 : viable cells without desiccation / exposure to vacuum, N : viable cells after desiccation / exposure to vacuum ($n = 3$). Black: Cells were desiccated on glass slides under anoxic conditions. Light grey: Cells were desiccated on quartz discs under anoxic conditions and exposed to vacuum (10^{-5} Pa) within the Trex-Box. Dark grey: Cells were desiccated on quartz discs under anoxic conditions and exposed to Martian atmosphere (Mars gas at a pressure of 10^{-3} Pa) within the Trex-Box.

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treatment. The survival rate of irradiated and desiccated *Y. intermedia* MASE-LG-1 shifted to lower survival values compared to cells irradiated in suspension which is likely caused by the effects of desiccation (Fig 5A, arrow marked with “desiccation”). The combination of desiccation and subsequent exposure to irradiation led to a higher survival rate than vice versa (Fig 5A). In a separate experiment, cells were desiccated under anoxic conditions and subsequently irradiated under oxic conditions to determine the effect of oxygen (Fig 5B). When the cells were exposed to oxygen during dry exposure to ionizing radiation the survival rate was lower than after anoxic exposure to ionizing radiation (Fig 5B, arrow marked with “oxygen”). This “oxygen” effect indicates that oxygen functioned as an additional stress parameter.

Nutrient limitation influences desiccation tolerance

The growth rate in optimal growth conditions was varying with the dilution factor of the medium. A 1:10 dilution of the MASE I medium did not cause a significant change in growth behavior (Fig 6A). Higher dilution (1:50) led to a reduction of the growth rate of one order of magnitude compared to 1:10 diluted medium (Fig 6A). The survival after one day of desiccation was independent of whether the medium was diluted 1:10 or 1:50. In contrast, compared to undiluted medium, the survival rate was significantly lower: S (undiluted, 24 h) = 6.7×10^{-2} vs. S (1:10, 24 h) = 3.7×10^{-5} (Fig 6A).

The radiation sensitivity slightly increased under nutrient limited conditions (1:50). Cells, grown under nutrient limitation (1:10) showed nearly the same tolerance to ionizing radiation

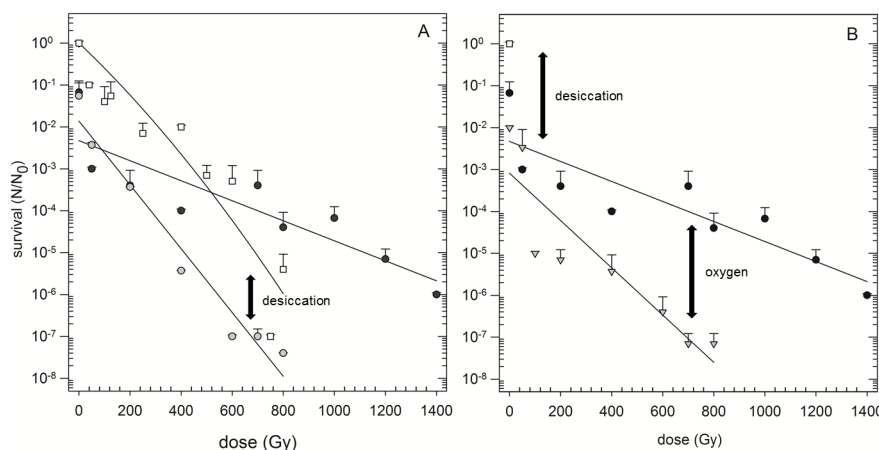


Fig 5. Survival after combined stresses (desiccation and radiation). Survival of *Y. intermedia* MASE-LG-1 after desiccation and irradiation in combination (A) and desiccation and irradiation in the presence of oxygen (B). N_0 : Viable cells after desiccation / irradiation. N : Viable cells after desiccation / irradiation ($n = 3$). (A) White squares: Cells were exposed to ionizing radiation under anoxic conditions in liquid culture medium. Black circles: Cells were desiccated (24 h) under anoxic conditions and subsequently exposed to ionizing radiation under anoxic conditions. Grey circles: Cells were exposed to ionizing radiation under anoxic conditions and subsequently desiccated (24 h) under anoxic conditions. (B) Black circles: Cells were desiccated (24 h) under anoxic conditions and subsequently exposed to ionizing radiation under anoxic conditions. White triangles: Cells were desiccated (24 h) under anoxic conditions and subsequently exposed to ionizing radiation under oxic conditions. White square: Survival of *Y. intermedia* MASE-LG-1 without desiccation and irradiation treatment.

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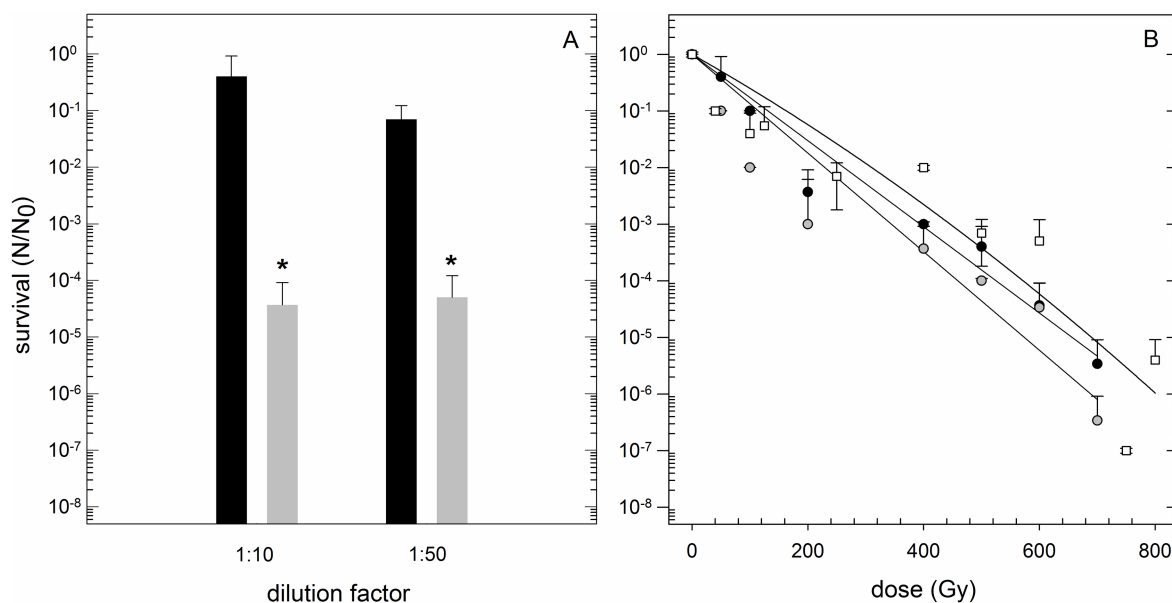


Fig 6. Influence of nutrient limitation. Influence of nutrient limitation on tolerance to desiccation (A) and ionizing radiation (B). N_0 : Viable cells without desiccation / irradiation, N : Viable cells after desiccation / irradiation. Recovery was performed under standard cultivation conditions ($n = 3$). (A) Black columns: growth in diluted medium after standard cultivation time (24 h). MASE I medium including all supplements was diluted 1:10 / 1:50 before inoculation. Grey columns: survival of *Y. intermedia* MASE-LG-1 in diluted medium (1:10 / 1:50) after desiccation (24 h) under anoxic conditions. Asterisks denote significant difference ($p < 0.05$) to the control (desiccation in full medium). (B) Cells grown under a limited set of nutrients were exposed to ionizing radiation up to 800 Gy under anoxic conditions. White squares: MASE I cultivation medium including all supplements without dilution. Black circles: MASE I cultivation medium, including all supplements was diluted 1:10 before inoculation. Grey circles: MASE I cultivation medium, including all supplements was diluted 1:50 before inoculation.

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as cells which were grown in undiluted medium (Fig 6B). This was quantified in the calculated D_{10} -values: D_{10} -value (1:10) 185 Gy, D_{10} -value (1:50) 175 Gy compared to the control (190 Gy).

Survival after exposure to low temperatures, salts and oxidizing compounds

Survival of *Y. intermedia* MASE-LG-1 at low temperature (-28°C , -86°C) was tested and the results revealed that the strain can survive low temperatures without added cryoprotectants. After 24 h at -28°C and -86°C no remarkable loss in survivability was observed. In contrast, after one day at $+90^{\circ}\text{C}$ no viable cells were detected (data not shown).

Table 2 summarizes the tolerance of the *Yersinia* strain to potentially cell damaging compounds. *Y. intermedia* MASE-LG-1 tolerated different salt concentrations and showed survival after 15 min exposure up to 6.0 M NaCl, 2.0 M CaCl_2 , and 3.0 M MgCl_2 . *Y. intermedia* MASE-LG-1 cultures were exposed to oxidizing compounds (hydrogen peroxide and perchlorates) by adding appropriate amounts of stock solutions of the corresponding reagent directly to the grown cultures. Fifteen minutes exposure of oxidizing compounds led to a change in survivability of the strain. For example, no viable cells were detected in hydrogen peroxide concentrations greater than 100 mM. In contrast to the sensitivity against H_2O_2 , *Y. intermedia* MASE-LG-1 was tolerant against different perchlorates. Since both salts, $\text{Mg}(\text{ClO}_4)_2$ and $\text{Ca}(\text{ClO}_4)_2$, are formed from divalent cations, the tolerance to ClO_4^- ions up to 2.4 M was comparable.

Perchlorates influence desiccation tolerance

Y. intermedia MASE-LG-1 showed a high tolerance against short term exposure to perchlorate ions (2.0–2.4 M; Table 2). Consequently, the combination of perchlorates (in lower concentrations) and subsequent desiccation was investigated. Perchlorates had a significant negative influence on the desiccation tolerance of *Y. intermedia* MASE-LG-1 (Fig 7A). In the presence of perchlorates, the survival rate was reduced five to six orders of magnitude than what was observed after desiccation for 24 h without perchlorates. The reduction in the survival rate after desiccation was the same for both 0.5% and 1.0% perchlorates (concentrations for perchlorates in % are w/v). Amongst the particular perchlorates used, NaClO_4 had the strongest effect on the desiccation tolerance of *Y. intermedia* MASE-LG-1.

There was no statistical difference in survival between irradiated cells with and without perchlorates and amongst the two concentrations of the tested perchlorates (0.5% and 1.0%). All regression lines clustered together in a bundle (Fig 7B, ANOVA p -value = 0.934). The similarities in the survival curves are also noticeable in the calculated D_{10} -values (Table 3).

Table 2. Overview of components and concentrations at which growth / survival was detectable after exposure of 15 minutes.

Compound	Concentration
NaCl	$\leq 6.0 \text{ M}$
CaCl_2	$\leq 2.0 \text{ M}$
MgCl_2	$\leq 3.0 \text{ M}$
H_2O_2	$\leq 100 \text{ mM}$
NaClO_4	$\leq 2.4 \text{ M}$
$\text{Mg}(\text{ClO}_4)_2$	$\leq 1.2 \text{ M}$
$\text{Ca}(\text{ClO}_4)_2$	$\leq 1.0 \text{ M}$

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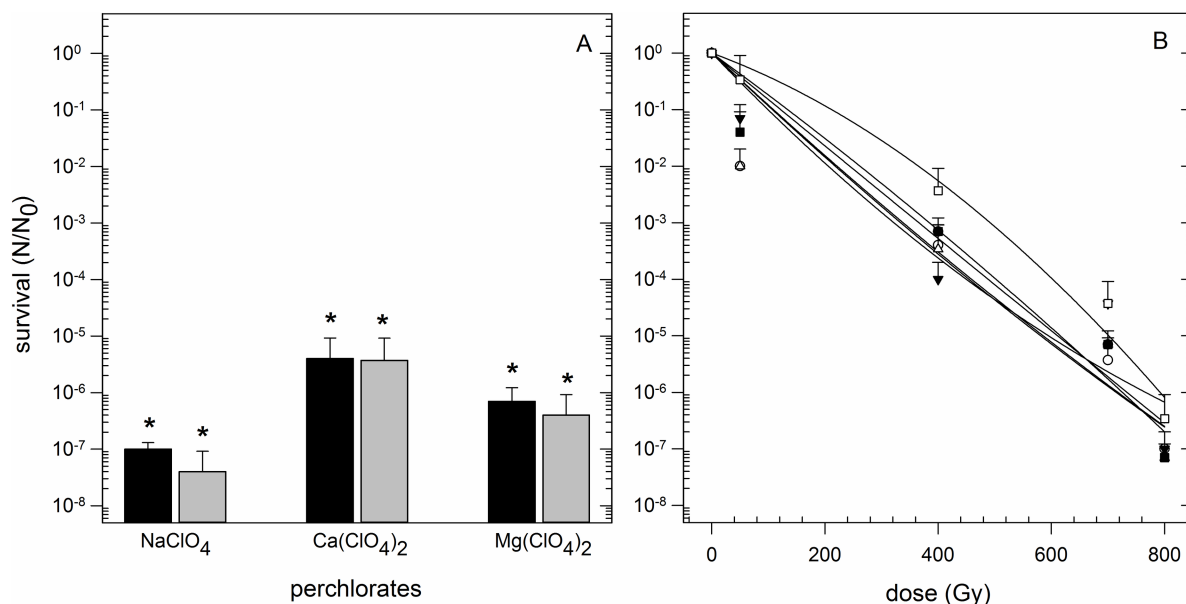


Fig 7. Influence of perchlorates. Influence of perchlorates on tolerance to desiccation (A) and ionizing radiation (B). N_0 : Viable cells without desiccation / irradiation, N : Viable cells after desiccation / irradiation. Recovery was performed under standard cultivation conditions without perchlorate ($n = 3$). Asterisks denote significant difference ($p < 0.05$) to the control (survival after desiccation without perchlorates). (A) Black columns: Cells were exposed (15 min) to 0.5% perchlorate (0.5% $\text{NaClO}_4 = 35.6$ mM; 0.5% $\text{Ca}(\text{ClO}_4)_2 = 20.9$ mM; 0.5% $\text{Mg}(\text{ClO}_4)_2 = 22.4$ mM) before desiccation treatment (24 h, anoxic conditions). Grey columns: Cells were exposed (15 min) to 1.0% perchlorate (1.0% $\text{NaClO}_4 = 71.2$ mM; 1.0% $\text{Ca}(\text{ClO}_4)_2 = 41.9$ mM; 1.0% $\text{Mg}(\text{ClO}_4)_2 = 44.8$ mM) before desiccation treatment (24 h, anoxic conditions). (B) Cells were exposed (15 min) to the indicated perchlorates before treatment with ionizing radiation up to 800 Gy. Black circles: 0.5% $\text{Mg}(\text{ClO}_4)_2$; White circle: 1% $\text{Mg}(\text{ClO}_4)_2$; Black triangle: 0.5% NaClO_4 ; White triangle: 1% NaClO_4 ; Black square: 0.5% $\text{Ca}(\text{ClO}_4)_2$; White square: 1% $\text{Ca}(\text{ClO}_4)_2$.

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Discussion

Mars is a hostile place for biological processes: anoxic atmosphere, low pressure, a low content of nutrients, presence of cell-damaging compounds, scarcity of liquid water, occurrence of solar and cosmic radiation, and extreme low temperature prevail [4]. Requirements for habitability and microbial life (as we know it), include the availability of traces of liquid water. The lowest water activity measured for organisms from Earth is approximately 0.61 [33]. Additionally, an energy source (chemical or solar), and chemical compounds suitable as nutrients are necessary [5, 34]. The presence of liquid water on the surface in Mars' early history is well-documented [1–3]. Although liquid water is less prevalent on the surface of Mars today, brines on

Table 3. Overview of calculated D_{10} -values of *Y. intermedia* MASE-LG-1 after exposure to ionizing radiation and its combination with other stresses.

Single stress test	Calculated D_{10} -value	Combined stress tests	Calculated D_{10} -values
X-rays	190 Gy	Nutrient limitation and X-rays 1:10	185 Gy
		Nutrient limitation and X-rays 1:50	175 Gy
		Perchlorate (0.5% ($\text{Mg}(\text{ClO}_4)_2$) and X-rays	173 Gy
		Perchlorate (1.0% ($\text{Mg}(\text{ClO}_4)_2$) and X-rays	129 Gy
		Perchlorate (0.5% (NaClO_4)) and X-rays	93 Gy
		Perchlorate (1.0% (NaClO_4)) and X-rays	125 Gy
		Perchlorate (0.5% ($\text{Ca}(\text{ClO}_4)_2$) and X-rays	183 Gy
		Perchlorate (1.0% ($\text{Ca}(\text{ClO}_4)_2$) and X-rays	189 Gy

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the surface and potentially in the subsurface is well-reported [35, 36]. Various chemical compounds which could function as redox couples can be found and could hypothetically be used by terrestrial organisms [37]. Also perchlorates and other oxidizing substances could be used under anoxic conditions to conserve energy for growth [38, 39]. Despite these factors that support habitability, Mars is also subject to many stresses that are potentially deleterious to life including desiccation, extremes of temperature and pH, ionizing radiation, oxidants and other potentially cell damaging compounds.

Although the response of a wide diversity of aerobic microorganisms has been studied under Martian stresses, systematic studies of anaerobic isolates are lacking. In particular, the response of anaerobic microorganisms to present-day Martian stresses has significant implications for planetary protection considerations which were established to counteract the deposition of terrestrial microbial life forms or signatures thereof onto the surface of Mars. This could be achieved by hardy microorganisms being able to survive the space travel or microorganisms that were accidentally deposited on Mars by robotic or human explorers with the potential to persist on the Martian surface. This process known as forward contamination might negatively impact life detection experiments [40]. We therefore require studies on the survival of cosmopolitan, anaerobic microorganisms that persist in many extreme environments on Earth to understand whether those same capabilities that allow for widespread persistence on Earth would allow for their persistence on Mars.

In this study, we isolated a species of *Yersinia* from an Icelandic environment and chose to use it as a model organism since it represents a highly adaptable, widespread genus found in many anoxic environments on Earth. Various *Y. intermedia* strains, the nearest relative to the isolated strain, as well as other *Yersinia* strains are found in freshwater or sewage all over the world [31]. All known strains of *Yersinia* are non-spore forming and for some, especially virulent strains, an ability to survive in aquatic environments and in humidified soils (up to 280 days) is already known [41–43]. It was shown that the virulent *Yersinia pestis* strain CO92 can persist after short term exposure (up to one hour) on different dry surfaces like glass and steel [44].

The persistence of *Yersinia* in many environments raises questions about its metabolic activity in those environments. Initially we attempted cultivation of enrichments at a range of pH conditions, but we were unable to isolate organisms at low pH and low temperatures. The optimal growth conditions of the isolated *Yersinia* strain and the environment bulk pH and temperature of the lake water are different. This discrepancy could be explained with the existence of microhabitats within the lake: e.g. at the bottom of the lake there are volcanic/hydrothermal activities probably due to hot springs [17]. Consequently, micro habitats / niches with optimal growth conditions for the strain could occur. Transcriptomic analyses for another *Y. intermedia* strain showed an adaptation mechanism for acid tolerance. This mechanism is especially active if the cells are growing under anoxic conditions [45]. Additionally, there is the discrepancy between the temperature we eventually used for cultivation (30°C) and the natural environment (4°C). The ability to accumulate protective cold shock proteins is known for the genus *Yersinia* [46]. Owing to their pathogenic lifestyle, the genus *Yersinia* is described as highly adaptable [47]. The hostile conditions within the natural environment (i.e. pH extremes, lack of oxygen, and starvation) could result in a pre-adaptation process such as genomic reorganization or even mutation as it is reported for *Escherichia coli* [48] and could lead to the high polyextremotolerance of the strain we isolated. Our findings do not allow us to determine unequivocally the activity of our isolate in the lake. However, its survival and maintenance of viability in such an extreme acidic and cold environment and the known widespread distribution of this anaerobic genus in diverse environments around the world make it an ideal organism with which to assess the capabilities of highly adaptable anaerobic organisms to

survive Mars-like conditions. Survival studies focused on Martian UV radiation have been conducted with other widespread distributed organisms like representatives from the genera *Deinococcus* or *Bacillus* [11, 49].

Y. intermedia MASE-LG-1 was found to be very desiccation tolerant. After 83 days of desiccation the survival rate was lowered only by four orders of magnitude. By comparison, *Hydrogenothermus marinus*, a desiccation-tolerant anaerobic organism also isolated from an aquatic system showed a reduction in its survival rate of 10^{-6} after 112 days of desiccation [28]. Compared with other desiccation-tolerant aerobic organisms, such as *Deinococcus radiodurans*, the survival rate after desiccation was lower. We showed that 0.37% of the *Yersinia* cells survived after a desiccation time of 14 days. The survival rate of *D. radiodurans* does not change within the first 100 hours of desiccation and after 400 hours (~ 16 days) 40% survived [50]. The exposure of dried cells to vacuum / Martian atmosphere had no effect on the survival; i.e. once the cells survived the first desiccation step, they were stable with respect to additional desiccation in the form of vacuum and Martian atmosphere. In contrast to our findings, an impact of space vacuum and Martian atmosphere on survival rate of *D. radiodurans* has been observed: after exposure to space vacuum (10^{-6} Pa, three days) the survival rate was reduced more than three orders of magnitude compared to desiccated cells [51]. However, if *D. radiodurans* is only exposed to Martian atmosphere (7 days) the survival rate was reduced less than one order of magnitude [52].

Research using *D. radiodurans*, the model organism in radiation microbiology, suggested that desiccation tolerance and radiation resistance / tolerance are correlated [53, 54]. It has been suggested that the extraordinary high radiation tolerance, which is typical for several *Deinococcus* strains, is a by-product of desiccation tolerance [55, 56]. Additionally, this correlation was shown for different members of the genus *Chroococcidiopsis* [57]. However, this trend is in contrast to the results obtained with *Y. intermedia* MASE-LG-1. The isolate was desiccation resistant as described above but was sensitive to ionizing radiation. This was shown in the survival rates as well as in the calculated D_{10} -values. For comparison, the D_{10} -value given in literature for other vegetative cells such as *E. coli* (250 Gy [58]) and *Bacillus pumilus* (210 Gy [59]) are higher than the D_{10} -value of our isolate (190 Gy). D_{10} -values of other *Yersinia* strains from literature are lower (e.g. *Y. enterocolitica* D_{10} -value 118 Gy [60]). Interestingly, there is a known dependency between temperature and resistance to radiation: the lower the temperatures are during irradiation treatment, the higher the survival rate is. The D_{10} -value for *Y. enterocolitica* exposed to ionizing radiation at 0°C is 190 Gy, which is the same D_{10} -value as measured for the strain in this study at room temperature. The D_{10} -value of *Y. enterocolitica* after irradiation treatment at -20°C is 380 Gy; for -76°C even 550 Gy [61].

When the cells were desiccated and subsequently irradiated, they resisted a higher dose of ionizing radiation than in liquid culture. This effect, also known for some halophilic Archaea [62], can be explained by the fact that during desiccation at room temperature, which is near to the optimal growth temperature (30°C), the cells can react actively to the process of desiccation. Intracellular protection mechanisms can take place, e.g. entering a kind of “dormant state” [63]. Furthermore, under desiccation, in conditions with low water availability, there may be lower concentrations of reactive oxygen species caused by interactions of water with ionizing radiation. Therefore, one explanation might be that pre-adaptation to desiccation primed the cells to be less sensitive to other harmful effects e.g. ionizing radiation.

Y. intermedia MASE-LG-1, having the ability to grow under oxic as well as anoxic conditions, should be capable of resisting the negative effects of an oxygen exposure. It is known that the presence of oxygen during irradiation intensifies the effects of radiation [64, 65]. The strain showed oxygen sensitivity when the irradiation was performed under oxic conditions. As the cells were irradiated in a desiccated form, radiolysis of water and the occurrence of

oxygen radicals should be negligible [66]. However, the desiccation step does not completely remove the intracellular H₂O. This residual water in combination with the applied radiation can lead to an intracellular radical formation by radiolysis. This may explain why the impact was increased with a greater applied dose. When the cells were first irradiated and then desiccated a typical additive effect was observed. The regression line was lowered by two orders of magnitude which was identical to one day of desiccation. The effects of the stresses accumulate additively and the change was the sum of the individual effects [67].

On Mars, only a limited set of nutrients is available and additional cell-damaging components such as high salt concentrations and perchlorates are present on its surface [30, 68, 69]. *Y. intermedia* MASE-LG-1 is on the one hand able to grow under a limited set of nutrients, but this nutrient deprivation influenced the desiccation tolerance in a negative way. Likewise, the strain tolerated a short-term exposure to high concentrations of NaCl and perchlorates, but these high salt concentrations and even Mars-like concentrations of perchlorate (0.5% / 1.0%) reduced survival after desiccation as it was already described for *H. marinus* [70]. The resistance of the *Yersinia* strain to hydrogen peroxide is lower than that reported for other hydrogen peroxide resistant aerobic organisms and in a comparable range as reported for hydrogen peroxide resistant anaerobic organisms like *Bacteroides fragilis* [71, 72]. *Y. intermedia* MASE-LG-1 shows poor survival under exposure to H₂O₂ as compared to aerobic *Acinetobacter gyllenbergii* 2P01AA, which is reported to show the highest known tolerance to hydrogen peroxide among Gram-negative and non-spore forming bacteria so far [71]. While *A. gyllenbergii* 2P01AA can survive an hour's exposure to 100 mM H₂O₂, after exposure to 320 mM, its survival was reduced by two orders of magnitude and no survival was measurable after exposure to 490 mM. *Y. intermedia* MASE-LG-1 can only survive for 15 min in 100 mM H₂O₂.

Y. intermedia MASE-LG-1 exhibits a high tolerance against short term exposure to high amounts of perchlorate ions (up to 2.4 M \pm 33.7%). There is little known about the upper survival limit under perchlorates exposure of other strains in literature. The cells are often only treated with low concentrations of perchlorate. For example, halophilic archaea are able to grow in 400 mM (\pm 5.6%) sodium perchlorate [73]. The methanogenic archaea *Methanothermobacter wolfeii*, and *Methanosarcina barkeri* were able to survive exposure to 5% and 25% perchlorate (magnesium perchlorate, calcium perchlorate, sodium perchlorate) for variable time periods with *M. wolfeii* surviving the 25% (\pm 1.5 M magnesium perchlorate) concentration for longer (72 hours) than *M. barkeri* [74].

When compared to real Martian conditions, *Y. intermedia* MASE-LG-1 was able to survive some of these Martian conditions for limited periods. On Mars, oxygen is at 0.15% in the thin atmosphere at a pressure of 6 mbar [3]. The *Y. intermedia* MASE-LG-1 was able to withstand Martian atmosphere desiccation for weeks. Low temperatures prevail on Mars' surface: the average surface temperature is -55°C and at regions at the Martian equator the temperatures sometimes rise above 0°C [3]. The strain was able to tolerate exposures to temperatures down to -86°C without loss in survivability. A putative role of intracellular accumulated cryoprotective compatible solutes cannot be excluded, which has also been proven for *Y. enterocolitica* [75]. The radiation dose rate of ionizing radiation on the surface of Mars was measured and calculated to be up to 0.21 mGy per day [76, 77]. Given the results of this study, *Y. intermedia* MASE-LG-1 might possibly survive the radiation doses occurring on the Martian surface or in the first centimeter in the Martian soil / regolith for several years or decades [78]. On Mars, other types of radiation occur in galactic cosmic rays and solar energetic particles, which we did not study here.

In conclusion, our data show that even a cosmopolitan, adaptable species of anaerobic microorganism from a genus widely distributed and persistent in many extreme environments on Earth ultimately succumbed to stresses associated with Martian surface conditions. Our

data suggest that many similar anaerobic microorganisms might well survive on Mars for short periods of time, but that ultimately they will be killed by Martian conditions. Our data provide a systematic investigation of an anaerobic model microorganism in a variety of Martian extremes. In Martian conditions under which many of these extremes are mitigated (such as subsurface environments) these types of persistent organisms may well have better chances to grow.

Supporting information

S1 Fig. Dendrogram *Y. intermedia* MASE-LG-1. Neighbor-joining tree based on 16S rDNA sequences showing the phylogenetic position of *Y. intermedia* MASE-LG-1 related to 14 species of the genus *Yersinia*. DNA sequences were aligned using BioEdit. The dendrogram was constructed by using MEGA6. Bar: 0.01 substitutions per nucleotide position. (TIF)

S1 Table. Results of the API R 20 A and Rapid ID 32 A tests. + *Y. intermedia* MASE-LG-1 metabolized the named substance; – *Y. intermedia* MASE-LG-1 did not metabolize the named substance. (DOCX)

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References

1. Pollack JB, Kasting JF, Richardson SM, Poliakov K. The case for a wet, warm climate on early Mars. *Icarus*. 1987; 71: 203–224. PMID: [11539035](https://pubmed.ncbi.nlm.nih.gov/11539035/)

2. Baker VR. Water and the Martian landscape. *Nature*. 2001; 412: 228–236. <https://doi.org/10.1038/35084172> PMID: 11449284
3. Jakosky BM, Phillips RJ. Mars' volatile and climate history. *Nature*. 2001; 412: 237–244. <https://doi.org/10.1038/35084184> PMID: 11449285
4. Horneck G. The microbial world and the case for Mars. *Planet Space Sci*. 2000; 48: 1053–1063.
5. Cockell CS, Bush T, Bryce C, Direito S, Fox-Powell M, Harrison JP et al. Habitability: a review. *Astrobiology*. 2016; 16: 89–117. <https://doi.org/10.1089/ast.2015.1295> PMID: 26741054
6. Wadsworth J, Cockell CS. Perchlorates on Mars enhance the bacteriocidal effects of UV light. *Sci Rep*. 2017; 7: 4662. <https://doi.org/10.1038/s41598-017-04910-3> PMID: 28684729
7. Cockell CS, Schuerger AC, Billi D, Friedmann EI, Panitz C. Effects of a simulated martian UV flux on the cyanobacterium, *Chroococcidiopsis* sp. 029. *Astrobiology*. 2005; 5: 127–140. <https://doi.org/10.1089/ast.2005.5.127> PMID: 15815164
8. Moeller R, Stackebrandt E, Reitz G, Berger T, Rettberg P, Doherty AJ, et al. Role of DNA repair by non-homologous-end joining in *Bacillus subtilis* spore resistance to extreme dryness, mono- and polychromatic UV, and ionizing radiation. *J Bacteriol*. 2007; 189: 3306–3311. <https://doi.org/10.1128/JB.00018-07> PMID: 17293412
9. Fendrihan S, Bércecs A, Lammer H, Musso M, Rontó G, Polacsek TK, et al. Investigating the effects of simulated Martian ultraviolet radiation on *Halococcus dombrowskii* and other extremely halophilic archaeobacteria. *Astrobiology*. 2009; 9: 104–112. <https://doi.org/10.1089/ast.2007.0234> PMID: 19215203
10. Wassmann M, Moeller R, Rabbow E, Panitz C, Horneck G, Reitz G, et al. Survival of spores of the UV-resistant *Bacillus subtilis* strain MW01 after exposure to low-Earth orbit and simulated Martian conditions: data from the space experiment ADAPT on EXPOSE-E. *Astrobiology*. 2012; 12: 498–507. <https://doi.org/10.1089/ast.2011.0772> PMID: 22680695
11. Newcombe DA, Schuerger AC, Benardini JN, Dickinson D, Tanner R, Venkateswaran K. Survival of spacecraft-associated microorganisms under simulated martian UV irradiation. *Appl Environ Microbiol*. 2005; 71: 8147–856. <https://doi.org/10.1128/AEM.71.12.8147-8156.2005> PMID: 16332797
12. Stieglmeier M, Wirth R, Kminek G, Moissl-Eichinger C. Cultivation of anaerobic and facultatively anaerobic bacteria from spacecraft-associated clean rooms. *Appl Environ Microbiol*. 2009; 75: 3484–3491. <https://doi.org/10.1128/AEM.02565-08> PMID: 19363082
13. Probst A, Vaishampayan P, Osman S, Moissl-Eichinger C, Andersen GL, Venkateswaran K. Diversity of anaerobic microbes in spacecraft assembly clean rooms. *Appl Environ Microbiol*. 2010; 76: 2837–2845. <https://doi.org/10.1128/AEM.02167-09> PMID: 20228115
14. La Duc MT, Dekas A, Osman S, Moissl C, Newcombe D, Venkateswaran K. Isolation and characterization of bacteria capable of tolerating the extreme conditions of clean room environments. *Appl Environ Microbiol*. 2007; 73:2600–2611. <https://doi.org/10.1128/AEM.03007-06> PMID: 17308177
15. DeVincenzi DL, Race MS, Klein HP. Planetary protection, sample return missions and Mars exploration: History, status, and future needs. *J Geophys Res*. 1998; 103: 28577–28585.
16. Cockell CS, Schwendner P, Perras A, Rettberg P, Beblo-Vranesovic K, Bohmeier M, et al. Anaerobic microorganisms in astrobiological analog environments: from field site to culture collection. *Int J Astrobiol*. 2017; 0: 1–15 (in press).
17. Thorarinsson S. Grænavatn and Gestsdavvatn. *Geografisk Tidsskrift*. 1953; 52.
18. Balch WE, Fox GE, Magrum LJ, Woese CR, Wolfe RS. Methanogens: reevaluation of a unique biological group. *Microbiol Rev*. 1979; 43: 260–296. PMID: 390357
19. Miller TL, Wolin MJ. A serum bottle modification of the hungate technique for cultivating obligate anaerobes. *Appl Microbiol*. 1974; 27: 985–987. PMID: 4598231
20. Tillett D, Neilan BA. Xanthogenate nucleic acid isolation from cultured and environmental cyanobacteria. *J Phycol*. 2000; 36: 251–258.
21. Burggraf S, Olsen GJ, Stetter KO, Woese CR. A phylogenetic analysis of *Aquifex pyrophilus*. *Sys Appl Microbiol*. 1992; 15: 352–356.
22. Lane DJ (1991) 16S/23S rRNA sequencing. In: Stackebrandt E, Goodfellow M, editors *Nucleic acid techniques in bacterial systematics*. John Wiley & Sons. Chichester, England. 1991. pp. 125–175.
23. Hall A. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symp Ser*. 1999; 41: 95–98.
24. Tamura K, Stecher G, Peterson D, Filipski A, Kumar S. MEGA6: molecular evolutionary genetics analysis version 6.0. *Mol Biol Evol*. 2013; 30: 2725–2729. <https://doi.org/10.1093/molbev/mst197> PMID: 24132122

25. Schumann P, Maier T. MALDI-TOF mass spectrometry applied to classification and identification of bacteria. *Method Microbiol.* 2014; 41: 275–306.
26. Franson MAH (ed.). Standard methods for the examination of water and wastewater. In: American Public Health Association, 16th edition, Washington DC; 1985.
27. Harm W. Biological effects of ultraviolet radiation. Cambridge University Press, Cambridge; 1980.
28. Beblo K, Rabbow E, Rachel R, Huber H, Rettberg P. Tolerance of thermophilic and hyperthermophilic microorganisms to desiccation. *Extremophiles.* 2009; 13: 521–531. <https://doi.org/10.1007/s00792-009-0239-1> PMID: 19333686
29. Beblo K, Douki T, Schmalz G, Rachel R, Wirth R, Huber H, et al. Survival of thermophilic and hyperthermophilic microorganisms after exposure to UV-C, ionizing radiation and desiccation. *Arch Microbiol.* 2011; 193: 797–809. <https://doi.org/10.1007/s00203-011-0718-5> PMID: 21638055
30. Kounaves SP, Chaniotakis NA, Chevrier VF, Carrier BL, Folds KE, Hansen VM, et al. Identification of the perchlorate parent salts at the Phoenix Mars landing site and possible implications. *Icarus.* 2014; 232: 226–231.
31. Brenner DJ, Bercovier H, Ursing J, Alonso MJ, Steigerwalt AG, Fanning GR, et al. *Yersinia intermedia*: a new species of Enterobacteriaceae composed of rhamnose-positive, melibiose-positive, raffinose-positive strains (formerly called *Yersinia enterocolitica* or *Yersinia enterocolitica*-like). *Curr Microbiol.* 1980; 4: 207–212.
32. Bercovier H, Steigerwalt AG, Guiry A, Huntley-Carter G, Brenner DJ. *Yersinia aldovae* (formerly *Yersinia enterocolitica*-like group X2): a new species of Enterobacteriaceae isolated from aquatic ecosystems. *Int J Syst Bacteriol.* 1984; 34: 166–172.
33. Stevenson A, Cray JA, Williams JP, Santos R, Sahay R, Neuenkirchen N, et al. Is there a common water-activity limit for the three domains of life? *ISME J.* 2015; 9: 1333–1351. <https://doi.org/10.1038/ismej.2014.219> PMID: 25500507
34. Moissl-Eichinger C, Cockell C, Rettberg P. Venturing into new realms? Microorganisms in space. *FEMS Microbiol Rev.* 2016; pii: fuw015.
35. Martín-Torres FJ, Zorzano MP, Valentín-Serrano P, Harri AM, Genzer M, Kempainen O, et al. Transient liquid water and water activity at Gale crater on Mars. *Nat Geosci.* 2015; 8: 357–361.
36. McEwen AS, Ojha L, Dundas CM, Mattson SS, Byrne S, Wray JJ, et al. Seasonal flows on warm Martian slopes. *Science.* 2011; 333: 740–743. <https://doi.org/10.1126/science.1204816> PMID: 21817049
37. Westall F, Loizeau D, Foucher F, Bost N, Bertrand M, Vago J, et al. Habitability on Mars from a microbial point of view. *Astrobiology.* 2013; 13: 887–897. <https://doi.org/10.1089/ast.2013.1000> PMID: 24015806
38. Hecht MH, Kounaves SP, Quinn RC, West SJ, Young SM, Ming DW, et al. Detection of perchlorate and the soluble chemistry of martian soil at the Phoenix lander site. *Science.* 2009; 325: 64–67. <https://doi.org/10.1126/science.1172466> PMID: 19574385
39. Liebensteiner MG, Stams AJ, Lomans BP. (Per)chlorate reduction at high temperature: physiological study of *Archaeoglobus fulgidus* and potential implications for novel souring mitigation strategies. *Int Biodeterior Biodegradation.* 2014; 96: 216–222.
40. Crawford RL. Microbial diversity and its relationship to planetary protection. *Appl Environ Microbiol* 2005; 71:4163–4168. <https://doi.org/10.1128/AEM.71.8.4163-4168.2005> PMID: 16085798
41. Agbalika F, Soltandallal M, Hartemann P. Isolation of *Yersinia* from surface water: comparative study of three selective media. *Water Res.* 1985; 19: 1255–1258.
42. Rose LJ, Donlan R, Banerjee SN, Arduino MJ. Survival of *Yersinia pestis* on environmental surfaces. *Appl Environ Microbiol.* 2003; 69: 2166–2171. <https://doi.org/10.1128/AEM.69.4.2166-2171.2003> PMID: 12676697
43. Ayyadurai S, Houhamdi L, Lepidi H, Nappez C, Raoult D, Drancourt M. Long-term persistence of virulent *Yersinia pestis* in soil. *Microbiology.* 2008; 154: 2865–2871. <https://doi.org/10.1099/mic.0.2007/016154-0> PMID: 18757820
44. Gut IM, Bartlett RA, Yeager JJ, Leroux B, Ratnesar-Shumate S, Dabisch P, et al. Extraction of aerosol-deposited *Yersinia pestis* from indoor surfaces to determine bacterial environmental decay. *Appl Environ Microbiol.* 2016; 82: 2809–2818. <https://doi.org/10.1128/AEM.03989-15> PMID: 26944839
45. Babujee L, Balakrishnan V, Kiley PJ, Glasner JD, Perna NT. Transcriptome changes associated with anaerobic growth in *Yersinia intermedia* (ATCC29909). *PloS One.* 2013; 8:e76567. <https://doi.org/10.1371/journal.pone.0076567> PMID: 24116118
46. Keto-Timonen R, Hietala N, Palonen E, Hakakorpi A, Lindström M, Korkeala H. Cold shock proteins: a minireview with special emphasis on Csp-family of enteropathogenic *Yersinia*. *Front Microbiol.* 2016; 7: 1151. <https://doi.org/10.3389/fmicb.2016.01151> PMID: 27499753

47. McNally A, Thomson NR, Reuter S, Wren BW. 'Add, stir and reduce': *Yersinia* spp. as model bacteria for pathogen evolution. *Nat Rev Microbiol*. 2016; 14: 177–190. <https://doi.org/10.1038/nrmicro.2015.29> PMID: 26876035
48. Jolivet-Gougeon A, David-Jobert S, Tamanai-Shacoori Z, Ménard C, Cormier M. Osmotic stress-induced genetic rearrangements in *Escherichia coli* H10407 detected by randomly amplified polymorphic DNA analysis. *Appl Environ Microbiol*. 2000; 66: 5484–5487. PMID: 11097933
49. Gómez F, Mateo-Martí E, Prieto-Ballesteros O, Martín-Gago J, Amils R. Protection of chemolithoautotrophic bacteria exposed to simulated Mars environmental conditions. *Icarus*. 2010; 209: 482–487.
50. Rettberg P, de La Vega U, Horneck G (2004) *Deinococcus radiodurans*—a model organism for life under Martian conditions. In: Harris RA, Ouwehand L, editors. *Proceedings of the Third European Workshop on Exo-Astrobiology*. 2004; 545: 59–62.
51. Saffary R, Nandakumar R, Spencer D, Robb FT, Davila JM, Swartz M, et al. Microbial survival of space vacuum and extreme ultraviolet irradiation: strain isolation and analysis during a rocket flight. *FEMS Microbiol Lett*. 2002; 215: 163–168. PMID: 12393217
52. Pogoda de La Vega U, Rettberg P, Reitz G. Simulation of the environmental climate conditions on Martian surface and its effect on *Deinococcus radiodurans*. *Adv Space Res*. 2007; 40: 1672–1677.
53. Mattimore V, Battista JR. Radioresistance of *Deinococcus radiodurans*: functions necessary to survive ionizing radiation are also necessary to survive prolonged desiccation. *J Bacteriol*. 1996; 178: 633–637. PMID: 8550493
54. Musilova M, Wright G, Ward JM, Dartnell LR. Isolation of radiation-resistant bacteria from Mars analog antarctic dry valleys by preselection, and the correlation between radiation and desiccation resistance. *Astrobiology*. 2015; 15: 1076–1090. <https://doi.org/10.1089/ast.2014.1278> PMID: 26684506
55. Ferreira AC, Nobre MF, Rainey FA, Silva MT, Wait R, Burghardt J, et al. *Deinococcus geothermalis* sp. nov. and *Deinococcus murrayi* sp. nov., two extremely radiation-resistant and slightly thermophilic species from hot springs. *Int J Syst Bacteriol*. 1997; 47: 939–947. <https://doi.org/10.1099/00207713-47-4-939> PMID: 9336890
56. De Groot A, Chapon V, Servant P, Christen R, Fischer-Le Saux M, Sommer S, et al. *Deinococcus deserti* sp. nov., a gamma-radiation-tolerant bacterium isolated from the Sahara Desert. *Int J Syst Evol Micr*. 2005; 55: 2441–2446.
57. Billi D, Friedmann EI, Hofer KG, Caiola MG, Ocampo-Friedmann R. Ionizing-radiation resistance in the desiccation-tolerant cyanobacterium *Chroococcidiopsis*. *Appl Environ Microbiol*. 2000; 66: 1489–1492. PMID: 10742231
58. Clavero MRS, Monk JD, Beuchat LR, Doyle MP, Brackett RE. Inactivation of *Escherichia coli* 0157:H7, *Salmonellae*, and *Campylobacter jejuni* in raw ground beef by gamma irradiation. *Appl Environ Microbiol*. 1994; 60: 2069–2075. PMID: 8031098
59. Parisi A, Antoine AD. Increased radiation resistance of vegetative *Bacillus pumilus*. *Appl Microbiol*. 1974; 28: 41–46. PMID: 4844266
60. El-Zawahry YA, Rowley DB. Radiation resistance and injury of *Yersinia enterocolitica*. *Appl Environ Microbiol*. 1979; 37: 50–54. PMID: 570017
61. Sommers CH, Niemira BA, Tunick M, Boyd G. Effect of temperature on the radiation resistance of virulent *Yersinia enterocolitica*. *Meat science*. 2002; 61: 323–328. PMID: 22060857
62. Leuko S, Domingos C, Parpart A, Reitz G, Rettberg P. The survival and resistance of *Halobacterium salinarum* NRC-1, *Halococcus hamelinensis*, and *Halococcus morrhuae* to simulated outer space solar radiation. *Astrobiology*. 2015; 15: 987–997. <https://doi.org/10.1089/ast.2015.1310> PMID: 26539978
63. Osborne DJ, Boubriak II. DNA and desiccation tolerance. *Seed Sci Res*. 1994; 4: 175–185.
64. Howard-Flanders P, Alper T. The sensitivity of microorganisms to irradiation under controlled gas conditions. *Rad Res*. 1957; 7: 518–540.
65. Schulte-Frohlind D, Ludwig DC, Rettberg P. Influence of thiols and oxygen on the survival of γ -irradiated plasmid DNA and cells. *Adv Space Res*. 1994; 14: 277–284.
66. Riley PA. Free radicals in biology: oxidative stress and the effects of ionizing radiation. *Int J Radiat Biol*. 1994; 65: 27–33. PMID: 7905906
67. Folt CL, Chen CY, Moore MV, Burnaford J. Synergism and antagonism among multiple stressors. *Limnol Oceanogr*. 1999; 44: 864–877.
68. Fisk MR, Giovannoni SJ. Sources of nutrients and energy for a deep biosphere on Mars. *J Geophys Res-Planet*. 1999; 104: 11805–11815.
69. Kerr RA. Pesky perchlorates all over Mars. *Science*. 2013; 340: 138.
70. Beblo-Vranesevic K, Huber H, Rettberg P. High tolerance of *Hydrogenothermus marinus* to sodium perchlorate. *Front Microbiol*. 2017; 8: 1369. <https://doi.org/10.3389/fmicb.2017.01369> PMID: 28769918

71. Derecho I, McCoy KB, Vaishampayan P, Venkateswaran K, Mogul R. Characterization of hydrogen peroxide-resistant *Acinetobacter* species isolated during the Mars Phoenix spacecraft assembly. *Astrobiology*. 2014; 14: 837–847. <https://doi.org/10.1089/ast.2014.1193> PMID: 25243569
72. Rocha ER, Selby T, Coleman JP, Smith CJ. Oxidative stress response in an anaerobe, *Bacteroides fragilis*: a role for catalase in protection against hydrogen peroxide. *J Bacteriol*. 1996; 178: 6895–6903. PMID: 8955312
73. Oren A, Bardavid RE, Mana L. Perchlorate and halophilic prokaryotes: implications for possible halophilic life on Mars. *Extremophiles*. 2014; 18: 75–80. <https://doi.org/10.1007/s00792-013-0594-9> PMID: 24150694
74. Kral TA, Goodhart TH, Harpool JD, Hearnberger CE, McCracken GL, McSpadden SW. Sensitivity and adaptability of methanogens to perchlorates: Implications for life on Mars. *Planet Space Sci*. 2016; 120: 87–95.
75. Park S, Smith LT, Smith GM. Role of glycine betaine and related osmolytes in osmotic stress adaptation in *Yersinia enterocolitica* ATCC 9610. *Appl Environ Microbiol*. 1995; 61: 4378–4381. PMID: 16535192
76. Matthiä D, Ehresmann B, Lohf H, Köhler J, Zeitlin C, Appel J, et al. The Martian surface radiation environment—a comparison of models and MSL/RAD measurements. *J Space Weather Space Clim*. 2016; 6: A13.
77. Hassler DM, Zeitlin C, Wimmer-Schweingruber RF, Ehresmann B, Rafkin S, Eigenbrode JL, et al. Mars' surface radiation environment measured with the Mars Science Laboratory's Curiosity rover. *Science*. 2014; 343: 1244797. <https://doi.org/10.1126/science.1244797> PMID: 24324275
78. Dartnell LR, Desorgher L, Ward JM, Coates AJ. Modelling the surface and subsurface martian radiation environment: implications for astrobiology. *Geophys Res Let*. 2007; 34.